Heredity **77** (1996) 665–668

Author index

Adams, W.T. *see* Burczyk, J.

Addis, G. *see* Marchi, A.

Allouis, S. *see* Higuet, D.

Andersson, S., Floral variation in *Saxifraga granulata*: phenotypic selection, quantitative genetics and predicted response to selection **217**

Apostolidis, A.P., Karakousis, Y., Triantaphyllidis, C., Genetic differentiation and phylogenetic relationships among Greek *Salmo trutta* L. (brown trout) populations as revealed by RFLP analysis of PCR amplified mitochondrial DNA segments **608**

Arnold, J. *see* Dean, R.

Baker, A.J. *see* Merilä, J.

Barilani, M. *see* Mantovani, B.

Barrett, J.A. *see* Ibrahim, K.M.

Barrett, S.C.H. *see* Eckert, C.G.

Bartos, P. *see* Hanusová, R.

Beall, E. *see* Morán, P.

Bergé, J. *see* Rooker, S.

Berlocher, S.H., McPheron, B.A., Population structure of *Rhagoletis pomonella*, the apple maggot fly **83**

Bertrand, D. *see* Hurst, G.D.D.

Beukeboom, L.W., Seif, M., Mettenmeyer, T., Plowman, A.B., Michiels, N.K., Paternal inheritance of B chromosomes in a parthenogenetic hermaphrodite **646**

Bezant, J., Laurie, D., Pratchett, N., Chojecki, J., Kearsey, M., Marker regression mapping of QTL controlling flowering time and plant height in a spring barley (*Hordeum vulgare* L.) cross **64**

Björklund, M. *see* Merilä, J.

Blackman, R.L., Spence, J.M., Field, L.M., Javed, N., Devine, G.J., Devonshire, A.L., Inheritance of the amplified esterase genes responsible for insecticide resistance in *Myzus persicae* (Homoptera: Aphididae) **154**

Blanco, G. *see* Sánchez, J.A.

Brown, J.K.M. *see* O'Hara, R.B.

Bryant, E.H., Meffert, L.M., Nonadditive genetic structuring of morphometric variation in relation to a population bottleneck **168**

Bubli, O.A., Rakitskaya, T.A., Imasheva, A.G., Variation of allozyme loci in populations of *Drosophila melanogaster* from the former USSR **638**

Bullock, D.W. *see* Williams, C.L.

Burczyk, J., Variance effective population size based on multilocus gamete frequencies in coniferous populations: an example of a Scots pine clonal seed orchard **74**

Burczyk, J., Adams, W.T., Shimizu, J.Y., Mating patterns and pollen dispersal in a natural knobcone pine (*Pinus attenuata* Lemmon.) stand **251**

Butlin, R.K., Co-ordination of the sexual signalling system and the genetic basis of differentiation between populations in the brown planthopper, *Nilaparvata lugens* **369**

Capula, M., Evolutionary genetics of the insular lacertid lizard *Podarcis tiliguerta*: genetic structure and population heterogeneity in a geographically fragmented species **518**

Carcereri, A. *see* Ennas, M.G.

Chapuisat, M. *see* Wedekind, C.

Chiang, C.-H. *see* Wang, C.-T.

Chojecki, J. *see* Bezant, J.

Christiansen, F.B. *see* Schierup, M.H.

Clabby, C. *see* Sánchez, J.A.

Clarke, R.T. *see* Raybould, A.F.

Collins, F.H. *see* Lehmann, T.

Comes, H.P., Kadereit, J.W., Genetic basis of speed of development in *Senecio vulgaris* L. var. *vulgaris*, *S. vulgaris* ssp. *denticulatus* (O.F. Muell.) P.D. Sell, and *Senecio vernalis* Waldst. & Kit. **544**

Courtois, B. *see* Huang, N.

Coutellec-Vreto, M.-A. *see* Guiller, A.

Crnjar, R. *see* Marchi, A.

Curtsinger, J.W. *see* Ross, M.J.

Dean, R., Arnold, J., Small sample properties for estimators of cytonuclear disequilibria **396**

Delesalle, V.A. *see* Mazer, S.J.

Devine, G.J. *see* Blackman, R.L.

Devonshire, A.L. *see* Blackman, R.L.

Dujardin, J.P. *see* Pereira, J.

Durand, P. *see* Trouvé , S.

Eckert, C.G., Manicacci, D., Barrett, S.C.H., Frequency-dependent selection on morph ratios in tristylous *Lythrum salicaria* (Lythraceae) **581**

Ennas, M.G., Suzuki, H., Menegazzi, M., Carcereri, A., Hanaoka, F., Gremo, F., Nieddu, M., Mezzanotte, R., On the presence of DNA polymerase α in human lymphocyte nuclei and chromosomes **186**

Exposito Hermosa, V. *see* Marchi, A.

Fanara, J.J., Hasson, E., Rodríguez, C., Santos, M., Fontdevila, A., The evolutionary history of *Drosophila buzzatii*. XXXIII. Are *Opuntia* hosts a selective factor for the inversion polymorphism? **500**

Field, L.M. *see* Blackman, R.L.

Flavin, F. see Sánchez, J.A.

Fontdevila, A. *see* Fanara, J.J.

Fontenille, D. *see* Lehmann, T.
Fuller, S.J., Mather, P.B., Wilson, J.C., Limited genetic differentiation among wild *Oryctolagus cuniculus* L. (rabbit) populations in arid eastern Australia **138**

García Guerreiro, M.P., Behaviour of the transposable elements *copia* and *mdg1* in hybrids between the sibling species *Drosophila melanogaster* and *D. simulans* **40**
García-Marín, J.L., Pla, C., Origins and relationships of native populations of *Salmo trutta* (brown trout) in Spain **313**
García-Vázquez, E. *see* Morán, P.
Goldson, S.L. *see* Williams, C.L.
Gourmet, C., Rayburn, A.L., Identification of RAPD markers associated with the presence of B chromosomes in maize **240**
Gremo, F. *see* Ennas, M.G.
Groeters, F.R., Maternally inherited sex ratio distortion as a result of a male-killing agent in *Spilostethus hospes* (Hemiptera: Lygaeidae) **201**
Guillemaud, T. *see* Rooker, S.
Guillemaud, T., Rooker, S., Pasteur, N., Raymond, M., Testing the unique amplification event and the worldwide migration hypothesis of insecticide resistance genes with sequence data **535**
Guiller, A., Coutellec-Vreto, M.-A., Madec, L., Genetic relationships among suspected contact zone populations of *Helix aspersa* (Gastropoda: Pulmonata) in Algeria **113**

Hager, B.J., Teale, S.A., The genetic control of pheromone production and response in the pine engraver beetle *Ips pini* **100**
Hammarton, T.C. *see* Hurst, G.D.D.
Hanaoka, F. *see* Ennas, M.G.
Hansen, M.M. *see* Nielsen, E.E.
Hanusová, R., Hsam, S.L.K., Bartos, P., Zeller, F.J., Suppression of powdery mildew resistance gene *Pm8* in *Triticum aestivum* L. (common wheat) cultivars carrying wheat–rye translocation T1BL .1RS **383**
Hasson, E. *see* Fanara, J.J.
Hawksworth, F.G. *see* Wiens, D.
Hawley, W.A. *see* Lehmann, T.
Hayter, J.B.R. *see* Ibrahim, K.M.
Hennon, P.E. *see* Wiens, D.
Hewitt, G.M. *see* Ibrahim, K.M.; Juan, C.
Higuet, D., Meráot, H., Allouis, S., Montchamp-Moreau, C., The relationship between structural variation and dysgenic properties of *P* elements in long-established *P*-transformed lines of *Drosophila simulans* **9**
Hilbish, T.J. *see* Rawson, P.D.
Hohls, T., Setting confidence limits to genetic parameters estimated by restricted maximum likelihood analysis of North Carolina design II experiments **476**
Holloway, G.J. *see* Ottenheim, M.M.
Hsam, S.L.K. *see* Hanusová, R.
Huang, N., Courtois, B., Khush, G.S., Lin, H., Wang, G., Wu, P., Zheng, K., Association of quantitative trait loci for plant height with major dwarfing genes in rice **130**
Humphreys, M.W., Pasakinskiene, I., Chromosome painting to locate genes for drought resistance transferred from *Festuca arundinacea* into *Lolium multiflorum* **530**
Hurst, G.D.D., Hammarton, T.C., Obrycki, J.J., Majerus, T.M.O., Walker, L.E., Bertrand, D., Majerus, M.E.N., Male-killing bacterium in a fifth ladybird beetle, *Coleomegilla maculata* (Coleoptera: Coccinellidae) **177**

Ibrahim, K.M. *see* Juan, C.
Ibrahim, K.M., Hayter, J.B.R., Barrett, J.A., Frequency changes in storage protein genes in a hybrid bulk population of barley **231**
Ibrahim, K.M., Nichols, R.A., Hewitt, G.M., Spatial patterns of genetic variation generated by different forms of dispersal during range expansion **282**
Imasheva, A.G. *see* Bubli, O.A.
Islam-Faridi, M.N., Worland, A.J., Law, C.N., Inhibition of ear-emergence time and sensitivity to day-length, determined by the group 6 chromosomes of wheat **572**

Javed, N. *see* Blackman, R.L.
Joshi, A., Knight, C.D., Mueller, L.D., Genetics of larval urea tolerance in *Drosophila melanogaster* **33**
Jourdane, J. *see* Trouvé, S.
Joyner, K.L. *see* Rawson, P.D.
Juan, C., Ibrahim, K.M., Oromi, P., Hewitt, G.M., Mitochondrial DNA sequence variation and phylogeography of *Pimelia* darkling beetles on the Island of Tenerife (Canary Islands) **589**

Kadereit, J.W. *see* Comes, H.P.
Kamau, L. *see* Lehmann, T.
Karakousis, Y. *see* Apostolidis, A.P.
Kearsey, M. *see* Bezant, J.
Khush, G.S. *see* Huang, N.
King, E.J. *see* Wiens, D.
Knight, C.D. *see* Joshi, A.

Lascoux, M. *see* Lee, J.K.
Laurie, D. *see* Bezant, J.
Law, C.N. *see* Islam-Faridi, M.N.
Lee, J.K., Lascoux, M., Nordheim, E.V., Number of lethal equivalents in human populations: how good are the previous estimates? **209**
Lefèvre, F. *see* Legionnet, A.
Legionnet, A., Lefevre, F., Genetic variation of the riparian pioneer tree species *Populus nigra* L. I. Study of population structure based on isozymes **629**
Lehmann, T., Hawley, W.A., Kamau, L., Fontenille, D., Simard, F., Collins, F.H., Genetic differentation of *Anopheles gambiae* populations from East and West Africa: comparison of microsatellite and allozyme loci **192**

Leonardi, S., Menozzi, P., Spatial structure of genetic variability in natural stands of *Fagus sylvatica* L. (beech) in Italy **359**
Lin, H. *see* Huang, N.
Lin, T.-P. *see* Wang, C.-T.
Loeschcke, V. *see* Nielsen, E.E.

Macas, E. *see* Wedekind, C.
MacNair, J. *see* Naylor, J.
McPheron, B.A. *see* Berlocher, S.H.
Madec, L. see Guiller, A.
Majerus, M.E.N. *see* Hurst, G.D.D.
Majerus, T.M.O. *see* Hurst, G.D.D.
Manicacci, D. *see* Eckert, C.G.
Mantovani, B., Tinti, F., Barilani, M., Scali, V., Current reproductive isolation between ancestors of natural hybrids in *Bacillus* stick insects (Insecta: Phasmatodea) **261**
Marchi, A., Addis, G., Exposito Hermosa, V., Crnjar, R., Genetic divergence and evolution of *Polyommatus coridon gennargenti* (Lepidoptera: Lycaenidae) in Sardinia **16**
Maside, X.R., Naveira, H.F., A polygenic basis of hybrid sterility may give rise to spurious localizations of major sterility factors **488**
Maside, X.R., Naveira, H.F., On the difficulties of discriminating between major and minor hybrid male sterility factors in *Drosophila* by examining the segregation ratio of sterile and fertile sons in backcrossing experiments **433**
Mather, P.B. *see* Fuller, S.J.
Mazer, S.J., Delesalle, V.A., Floral trait variation in *Spergularia marina* (Caryophyllaceae): ontogenetic, maternal family, and population effects **269**
Meetze, K. *see* Rawson, P.D.
Meffert, L.M. *see* Bryant, E.H.
Megard, R.O. *see* Ross, M.J.
Menegazzi, M. *see* Ennas, M.G.
Menozzi, P. *see* Leonardi, S.
Merçot, H. *see* Higuet, D.
Merilä, J., Bjîrklund, M., Baker, A.J., The successful founder: genetics of introduced *Carduelis chloris* (greenfinch) populations in New Zealand **410**
Mettenmeyer, T. *see* Beukeboom, L.W.
Mezzanotte, R. *see* Ennas, M.G.
Michiels, N.K. *see* Beukeboom, L.W.
Mogg, R.J. *see* Raybould, A.F.
Montchamp-Moreau, C. *see* Higuet, D.
Morán, P., Pendás, A.M., Beall, E., García-Vázquez, E., Genetic assessment of the reproductive success of Atlantic salmon precocious parr by means of VNTR loci **655**
Mueller, D.L. *see* Joshi, A.

Naveira, H.F. *see* Maside, X.R.
Naylor, J., Macnair, M.R., Williams, E.N.D., Poultons, P.R., A polymorhpism for phosphate uptake/arsenate tolerance in *Holcus lanatus* L.: is there a correlation with edaphic or environmental factors? **509**

Nichols, R.A. *see* Ibrahim, K.M.
Nickrent, D.L. *see* Wiens, D.
Nieddu, M. *see* Ennas, M.G.
Nielsen, E.E., Hansen, M.M., Loeschcke, V., Genetic structure of European populations of *Salmo salar* L. (Atlantic salmon) inferred from mitochondrial DNA **351**
Nordheim, E.V. *see* Lee, J.K.

Obrycki, J.J. *see* Hurst, G.D.D.
O'Donald, P. see Tomlinson, I.P.M.
O'Hara, R.B., Brown, J.K.M., Frequency- and density- dependent selection in wheat powdery mildew **439**
O'Malley, D.M. *see* Plomion, C.
Oromi, P. *see* Juan, C.
Ottenheim, M.M., Volmer, A.D., Holloway, G.J., The genetics of phenotypic plasticity in adult abdominal colour pattern of *Eristalis arbustorum* (Diptera: Syrphidae) **493**

Parker, P.G. *see* Peare, T.
Pasakinskiene, I. *see* Humphreys, M.W.
Pasteur, N. *see* Guillemaud, T.; Rooker, S.
Peare, T., Parker, P.G., Local genetic structure within two rookeries of *Chelonia mydas* (the Green turtle) **619**
Pendás, A.M. *see* Morán, P.
Pereira, J., Dujardin, J.P., Salvatella, R., Tibayrenc, M., Enzymatic variability and phylogentic relatedness among *Triatoma infestans*, *T. platensis*, *T. delpontei* and *T. rubrovaria* **47**
Pigliucci, M., Modelling phenotypic plasticity. II. Do genetic correlations matter? **453**
Pla, C. *see* García-Marín, J.L.
Plomion, C., O'Malley, D.M., Recombination rate differences for pollen parents and seed parents in *Pinus pinaster* **341**
Plowman, A.B. *see* Beukeboom, L.W.
Poulton, P.R. *see* Naylor, J.
Powell, R. *see* Sánchez, J.A.
Pratchett, N. *see* Bezant, J.
Pugh, A.R.G., Ritchie, M.G., Polygenic control of a mating signal in *Drosophila* **378**

Rakitskaya, T.A. *see* Bubli, O.A.
Ramos, D. *see* Sánchez, J.A.
Rasmuson, M., A case of abnormal sex-diamorphism for bristle number in *Drosophila melanogaster* **146**
Rawson, P.D., Joyner, K.L., Meetze, K., Hilbish, T.J., Evidence for intragenic recombination within a novel genetic marker that distinguishes mussels in the *Mytilus edulis* species complex **599**
Raybould, A.F., Mogg, R.J., Clarke, R.T., The genetic structure of *Beta vulgaris* ssp. *maritima* (sea beet) populations: RFLPs and isoenzymes show different patterns of gene flow **245**
Rayburn, A.L. *see* Gourmet, C.
Raymond, M. *see* Guillemaud, T.; Rooker, S.
Renaud, F. *see* Trouvé, S.

Ritchie, M.G. *see* Pugh, A.R.G.
Rodríguez, C. *see* Fanara, J.J.
Rooker, S. *see* Guillemaud, T.
Rooker, S., Guillemaud, T., Bergé, J., Pasteur, N., Raymond, M., Coamplification of esterase A and B genes as a single unit in *Culex pipiens* mosquitoes **555**
Ross, M.J., Curtsinger, J.W., Megard, R.O., Development of population structure in *Daphnia* clones in a stratified lake **292**
Rülicke, T. *see* Wedekind, C.

Salvatella, R. *see* Pereira, J.
Sánchez, J.A., Clabby, C., Ramos, D., Blanco, G., Flavin, F., Vazquez, E., Powell, R., Protein and microsatellite single locus variability in *Salmo salar* L. (Atlantic salmon) **423**
Santos, M. *see* Fanara, J.J.
Scali, V. *see* Mantovani, B.
Schierup, M.H., Christiansen, F.B., Inbreeding depression and outbreeding depression in plants **461**
Seif, M. see Beukeboom, L.W.
Shaw, C.G., III *see* Wiens, D.
Shimizu, J.Y. *see* Burczyk, J.
Simard, F. *see* Lehmann, T.
Spence, J.M. *see* Blackman, R.L.
Suzuki, H. *see* Ennas, M.G.

Tachida, H., Yoshimaru, H., Genetic diversity in partially selfing populations with the stepping-stone structure **469**
Teale, S.A. *see* Hager, B.J.
Thompson, R. *see* Whittaker, J.C.
Tibayrenc, M. *see* Pereira, J.
Tinti, F. *see* Mantovani, B.
Tomlinson, I.P.M., O'Donald, P., The influence of female viability differences on the evolution of mate choice **303**
Triantaphyllids, C. *see* Apostolidis, A.P.
Trouvé, S., Renaud, F., Durand, P., Jourdane, J., Selfing and outcrossing in a parasitic hermaphrodite helminth (Trematoda, Echinostomatidae) **1**

Vázquez, E. *see* Sánchez, J.A.
Visscher, P.M. *see* Whittaker, J.C.
Volmer, A.D. *see* Ottenheim, M.M.

Walker, L.E. *see* Hurst, G.D.D.
Wang, C.-T., Wang, W.-Y., Chiang, C.-H., Wang, Y-N., Lin, T.-P., Low genetic variation *in Amentotaxus formosana* Li. revealed by isozyme analysis and random amplified polymorphic DNA markers **388**
Wang, G. *see* Huang, N.
Wang, W.-Y. *see* Wang, C.-T.
Wang, Y.-N. *see* Wang, C.-T.
Wedekind, C., Chapuisat, M., Macas, E., Rülicke, T., Non-random fertilization in mice correlates with the MHC and something else **400**
Whittaker, J.C., Thompson, R., Visscher, P.M., On the mapping of QTL by regression of phenotype on marker-type **23**
Wiens, D., Nickrent, D.L., Shaw, C.G., III, Hawksworth, F.G., Hennon, P.E., King, E.J., Embryonic and host-associated skewed adult sex ratios in dwarf mistletoe **55**
Williams, C.L., Goldson, S.L., Bullock, D.W., Unusual mitochondrial genome in introduced and native populations of *Listronotus bonariensis* (Kuschel) **565**
Williams, E.N.D. *see* Naylor, J.
Wilson, I.F., Application of ecological genetics techniques to test for selection by habitat on allozymes in *Cepaea nemoralis* (L.) **324**
Wilson, J.C. *see* Fuller, S.J.
Worland, A.J. *see* Islam-Faridi, M.N.
Wu, P. *see* Huang, N.

Yoshimaru, H. *see* Tachida, H.

Zeller, F.J. *see* Hanusová, R.
Zheng, K. *see* Huang, N.

Heredity **77** (1996) 669–671

Keyword index

allozymes **83, 113, 138, 192, 324, 359, 423, 518, 638**
Amentotaxus formosana **388**
Anopheles gambiae **192**
arsenate tolerance **509**
asexuality **646**
assortative mating **1**
autogamy **269**

B chromosomes **240, 646**
balancing selection **245**
barley **231**
beetle **589**
biparental inbreeding **469**
bottlenecks **168, 410**
brown trout **313**

cactus hosts **500**
canalization **269**
Canary Islands **589**
Carduelis chloris **410**
Caryophyllaceae **269**
cell cycle **186**
Cepaea nemoralis **324**
Chagas disease **47**
Chelonia mydas **619**
chromatin **186**
chromosome 2 **530**
chromosome assay **146**
chromosomes **186**
Coleomegilla maculata **177**
colonization **282**
communication systems **100**
comparative mapping **64**
composite cross **231**
computer simulation **476**
consanguineous marriages **209**
contact zone **113**
control region **138**
courtship song **378**
Culex pipiens **535**
cytochrome oxidase I **589**
cytonuclear disequilibria **396**

Daphnia pulicaria **292**
day-length **572**
deleterious alleles **209**
density-dependent selection **33, 439**
dispersal **282**
DNA fingerprinting **619**
DNA polymerase α **186**
DNA synthesis **186**
Drosophila **378, 433, 488**
D. buzzatii **500**
Drosophila melanogaster **33, 40, 146, 638**
Drosophila simulans **9, 40**
drought resistance **530**
dwarf mistletoes **55**

ear-emergence time **572**
early flowering mutants **544**
Echinostoma caproni **1**
ecological genetics **324**
effective migration rate **351**
effective population size **655**
electrophoresis **16, 292**
epistasis **168, 461**
Eristalis **493**
error rates **476**
Erysiphe graminis f. sp. *tritici* **439**
esterase genes **154, 535, 555**
evolution **313, 535, 555**

Fagus sylvatica **359**
female choice **303**
female preference **369**
female-biased sex ratio **177**
female viability **303**
fertilization **400**
FISH **154**
fixation index **469**
floral morphology **217**
flowering inhibition **572**
flowering plants **55**
flowering time **64**
forest-tree populations **359**
founder events **282**
frequency-dependent selection **439, 581**

gamete choice **400**
gene amplification **154, 555**
gene flow **192, 245, 251, 282, 351, 518**
genetic basis **544**
genetic correlation **217, 453**
genetic differentiation **16, 138, 351, 518**
genetic distance **47**
genetic diversity **629**

genetic markers **1, 23**
genetic resources **629**
genetic structure **113, 245, 359, 619**
genetic variance components **476**
genetic variation **388, 469, 493, 565, 608**
genome evolution **261**
genomic *in situ* hybridization **530**
genomic map **341**
genotype–environment interactions **453**
geographical isolation **16**
geographical differentiation **469**
geographical variation **638**
Greece **608**
group 6 chromosomes **572**
gymnosperm **341**

habitat selection **500**
height **64**
Helix aspera **113**
heritability **100, 217**
heteroduplex analysis **138**
heterostyly **581**
heterozygosity **410**
Holcus lanatus L. **509**
hordein **231**
Hordeum vulgare **64**
horizontal transmission **201**
hoverflies **493**
hybrid males **433, 488**
hybrid zone **396**
hybridization **261**
hybridogenesis **261**
hydroacoustic **292**

immunoassay **154**
inbred lines **23**
inbreeding coefficient **209**
inherited symbiont **177**
insecticide resistance **154**
interpulse interval **378**
interspecific hybrids **40**
interval mapping **23**
intragenic recombination **599**
introduced species **410, 565**
introgression lines **530**
inversion polymorphism **500**
Ips pini **100**
ipsdienol **100**
isozymes **47, 74, 245, 313, 388, 629**

Lacertidae **518**
larval crowding **33**
lethal equivalents **209**
life history trait **544**
linkage disequilibria **396**
Listronotus bonariensis **565**
log-linear modelling **292**
Lolium–Festuca **530**
Lythrum salicaria **581**

Maghreb **113**
major genes **433, 488**
male-killing bacteria **177, 201**
Mantel test **245**
marginalized methods of moments **209**
maternal effects **269**
maternal inheritance **201**
mating signal **369**
mating success **251**
mating system **74, 251**
mating system evolution **461**
MHC **400**
microsatellite variation **192, 423**
minisatellite loci **655**
mitochondrial DNA **138, 351, 589, 608**
mitochondrial genome **565**
molecular markers **130**
multiple fertilizations **1**
Mytilus **599**

narrow-sense heritability **476**
natural populations **638**
natural selection **217, 324, 500**
neighbourhood model **251**
neutral theory **324**
nucleotide polymorphism **535**
number of genes **369**

optimal outcrossing **461**
Oryza sativa **130**
outcrossing **251**

P element **9**
parthenogenesis **261**
paternal inheritance **646**
paternity assessment **655**
phenotypic plasticity **453**
pheromones **100**
philopatry **619**
phosphate uptake **509**
phylogeny **608**
phylogeography **589**
Pinus attenuata **251**
P. pinaster **341**
P. sylvestris **74**
planarian **646**
plant sexual polymorphisms **581**
plasticity **493**

© The Genetical Society of Great Britain, *Heredity*, 77, 669–671.

Pm8 suppressor gene **383**
Podarcis tiliguerta **518**
pollen gamete genotypes **74**
polygenes **433, 488**
polymorphism **240, 509**
Polyommatus coridon **16**
polyphenolic adhesive protein **599**
population genetic structure **192**
population genetics **83, 192, 423, 453, 599**
population heterogeneity **518**
population size **581**
population structure **83, 282, 292, 629**
Populus nigra **629**
position effect **154**
postglacial expansion **113**
powdery mildew **231, 383**
precocious parr **655**
pseudogamous parthenogenesis **646**

quantitative genetics **168, 453, 476**
quantitative trait loci (QTL) **23, 64, 130, 369**

RAPDs **240, 341, 388**
reaction norm **493**
recombination **433**
recombination mapping **488**
recombination rate **341**
regression **23**
reproductive success **655**
resistance **555**
RFLPs **130, 245, 351, 565, 608**
RFLP-mapping **64**
Rhagoletis **83**

Salmo salar **351, 423, 655**
S. trutta **313, 608**
Saxifraga granulata **217**
Secale cereale **383**
second meiotic division **400**
seed orchard **74**
selection **231**
selfing **1, 469**
selfish DNA **646**
Senecio vernalis **544**
S. vulgaris **544**
sequences **555**
sex allocation **269**
sex ratio distortion **201**
sex ratios **55**
sex **341**
sex-dimorphism **146**
sexual selection, 303, 369 **400**
shell phenotypes **324**
simulation **282, 461**
spatial autocorrelation **359**
speciation **16, 369, 378, 589**
speed of development **544**
Spergularia **269**
sperm exchanges **1**
Spilostethus hospes **201**
statistical software procedures **476**
stepping-stone model **469**
sterility **433, 488**
sternopleural bristle number **146**
stress resistance **33**
sympatric speciation **83**

taxonomy **589**
temperature gradient gel electrophoresis **138**
Tephritidae **83**
transposable elements **40**
Triatominae **47**
tristyly **581**
Triticum aestivum **383**

underdominance **461**
unisexual evolution **261**
urea tolerance **33**
Uruguay **47**

variance effective number **74**
vernalization **572**
Viscum **55**

wheat powdery mildew **439**
wheat **572**
wheat–rye translocation **383**

X-linkage **33**

Zea mays **240**

© The Genetical Society of Great Britain, *Heredity*, **77**, 669–671.